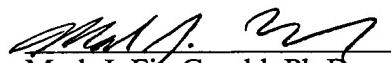


the figures directed herein as a result of the omission of Figure 10 from the application as filed. The amendments add no new matter.

The proposed amendments to the figures add a number to Figure 9 and re-number figures 11, 12 and 13 as filed to be numbers 10, 11 and 12, respectively, to correct the numbering after removal of reference to omitted Figure 10. Sheets with the proposed amendments to the figures marked in red, as well as clean sheets with the amendments are enclosed herein. Applicants submit that the amendments add no new matter.

Respectfully submitted,

10/12/01
Date


Mark J. Fitzgerald, Ph.D.
Registration No. 45,928
PALMER & DODGE LLP
One Beacon Street
Boston, MA 02108
Telephone: (617) 753-0451
Facsimile: (617) 227-4420

Version of Amendments Marked to Show Changes:

On page 1:

This application claims the priority [to] of U.S. Provisional application No.
[60/303,858, filed [7/9/01] July 9, 2001.]

- On page 19, delete paragraph 1, which starts "Figure 10 shows....":

[Figure 10 shows a partial chromatogram of the fifth step of purification of TIG2 from ascitic fluid. The active fractions (eluted with approximately 28% CH₃CN) of the previous step were diluted 6 fold with 0.1% TFA in H₂O and directly loaded onto a C18 reverse phase column (1mm x 50 mm, Vydac) pre-equilibrated with 5% CH₃CN/0.1% TFA in H₂O at a flow-rate of 0.1 ml/min. at room temperature. A 5-95% gradient of CH₃CN in 0.1%TFA was applied with a 0.3%/min slope between 25 and 45%. The activity was eluted at 40% CH₃CN (indicated by the black horizontal line).]

- On page 19, replace paragraphs 2, 3 and 4 and with the following replacement paragraphs 2, 3 and 4:

Figure [11] 10 shows the identification of a specific response for ChemR23 following screening of HPLC fractions obtained from the fractionation of human ovary ascites. The different fractions obtained following fractionation of human ovary ascites were diluted fivefold in the assay buffer and tested in an aequorin assay using a cell line expressing ChemR23 (open circles) or cell lines expressing unrelated receptors (closed triangles and squares). The response obtained for each fraction was normalized using the ATP response of each cell line.

Figure [12] 11 shows the activation of ChemR23 by conditioned medium of 293T cells transiently transfected with TIG2. 293T cells were transiently transfected with pCDNA3-TIG2 or with pCDNA3 alone (mock transfected). Increasing volumes of the supernatant collected 4 days after transfection were analyzed using a Microlumat in an aequorin-based assay with CHO cells expressing ChemR23. The assay was performed in triplicate, and SD is indicated. A representative experiment is shown.

Figure [13] 12 shows the characterization of antibodies directed against ChemR23 by flow cytometry.

- On page 54, replace the paragraph at lines 9-13 and with the following replacement paragraph:

The conditioned medium of COS-7, CHO-K1 and HEK 293 cells transfected with pCDNA3 encoding TIG2 was collected and used for aequorin assays on CHO cells expressing ChemR23. Results are shown in Figure [12] 11. Increasing amounts of conditioned supernatant resulted in an increase in luminescence in aequorin system cells expressing ChemR23.

- Replace the paragraph on page 54, line 25 to page 55, line 7 with the following replacement paragraph:

Figure [13] 12 shows the results of experiments to characterize the antibodies raised against ChemR23. A mixture of recombinant cells made up of 2/3 recombinant ChemR23 CHO cells and 1/3 mock-transfected CHO cells (negative control) was reacted with either a supernatant of cells expressing the anti ChemR23 5C 1H2 monoclonal antibody (thick line) or a supernatant from cells with no known antibody activity (thin line, grey filling). After staining with FITC labeled anti mouse Ig these preparations were analyzed by flow cytofluorometry. Results are displayed as a histogram of the number of cells (Events axis) expressing a given fluorescence (FL1-H axis). Monoclonal 5C 1H2 allowed the discrimination of the ChemR23 recombinant sub-population of cells from the negative control cells, as evidenced by the relative proportions of both types of cells. The background fluorescence of the assay is given by the second staining (grey filling).--